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(54) Title: ENDODERM, CARDIAC AND NEURAL INDUCING FACTORS (57) Abstract Novel proteins have been designated "cerberus" and "frzb-1", respectively. Cerberus is expressed as a secreted peptide during embryogenesis of the Xenopus embryo, and is expressed specifically in the head organizer region. This new molecule has endodermal, cardiac, and neural tissue inducing activity, that should prove useful in therapeutic, diagnostic, and clinical applications requiring regeneration, differentiation, or repair of these and other tissues. Frzb-1 is a soluble antagonist of growth factors of the Wnt family that acts by binding to Wnt growth factors in the extracellular space. A third novel protein is termed PAPC which promotes the formation of dorsal mesoderm and somites in the embryo.		

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ENDODERM, CARDIAC AND
NEURAL INDUCING FACTORS

5 Field of the Invention

 The invention generally relates to growth factors, neurotrophic factors, and their inhibitors, and more particularly to several new growth factors with neural, endodermal, and cardiac tissue inducing activity, to complexes and compositions including the factors, and to DNA or RNA coding sequences for the factors. Further, one of the novel growth factors should be useful in tumor suppression gene therapy.

 This application claims the benefit of U.S. Provisional Application No. 60/020,150, filed June 20, 1996.

 This invention was made with Government support under grant contract number HD-21502, awarded by the National Institutes of Health. The Government has

certain rights in this invention.

Background of the Invention

 Growth factors are substances, such as polypeptide hormones, which affect the growth of defined populations of animal cells *in vivo* or *in vitro*, but which are not nutrient substances. Proteins involved in the growth and differentiation of tissues may promote or inhibit growth, and promote or inhibit differentiation, and thus the general term "growth factor" includes cytokines, trophic factors, and their inhibitors.

Widespread neuronal cell death accompanies normal development of the central and peripheral nervous systems. Studies of peripheral target tissues during development have shown that neuronal cell death results from the competition among neurons for limiting amounts of survivor factors ("neurotrophic factors"). The earliest identified of these, nerve growth factor ("NGF"), is the most fully characterized and has been shown to be essential for the survival of sympathetic and neural crest-derived sensory neurons during early development of both chick and rat.

One family of neurotropic factors are the Wnts, which have dorsal axis-inducing activity. Most of the Wnt proteins are bound to cell surfaces. (See, e.g., Sokol et al., *Science*, 249, pp. 561-564, 1990.) Dorsal axis-inducing activity in *Xenopus* embryos by one member of this family (Xwnt-8) was described by Smith and Harland in 1991, *Cell*, 67, pp. 753-765. The authors described using RNA injections as a strategy for identifying endogenous RNAs involved in dorsal patterning to rescue dorsal development in embryos that were ventralized by UV irradiation.

Another member of the growth and neurotropic factor family was subsequently discovered and described by Harland and Smith, which they termed "noggin." (*Cell*, 70, pp. 829-840 (1992).) Noggin is a good candidate to function as a signaling molecule in Nieuwkoop's center, by virtue of its maternal transcripts, and in Spemann's organizer, through its zygotic organizer-specific expression. Besides noggin, other secreted factors may be involved in the organizer phenomenon.

Another *Xenopus* gene designated "chordin" that begins to be expressed in Spemann's organizer and that can completely rescue axial development in ventralized

embryos was described by Sasai et al., *Cell*, 79, pp. 779-790, 1994. In addition to dorsalizing mesoderm, chordin has the ability to induce neural tissue and its activities are antagonized by Bone Morphogenetic Protein-4 (Sasai et al., *Nature*, 376, pp. 333-336, 1995).

Therefore, the dorsal lip or Spemann's organizer of the *Xenopus* embryo is an ideal tissue for seeking novel growth and neurotrophic factors. New growth and neurotrophic factors are useful agents, particularly those that are secreted due to their ability to be used in physiologically active, soluble forms because these factors, their receptors, and DNA or RNA coding sequences therefore and fragments thereof are useful in a number of therapeutic, clinical, research, diagnostic, and drug design applications.

Summary of the Invention

In one aspect of the present invention, the sequence of the novel peptide that can be in substantially purified form is shown by SEQ ID NO:1. The *Xenopus* derived SEQ ID NO:1 has been designated "cerberus," and this peptide is capable of inducing endodermal, cardiac, and neural tissue development in vertebrates when expressed. The nucleotide sequence which, when expressed results in cerberus, is illustrated by SEQ ID NO:2. Since peptides of the invention induce endodermal, cardiac, and neural tissue differentiation in vertebrates, they should be able to be prepared in physiologically active form for a number of therapeutic, clinical, and diagnostic applications.

Cerberus was isolated during a search for molecules expressed specifically in Spemann's organizer containing a secretory signal sequence. In addition to cerberus, two other novel cDNAs were identified.

The *Xenopus* derived peptide that can be deduced from SEQ ID NO:3 encodes a novel protein we had earlier designated as "frazzled," a secreted protein of 318 amino acids that has dorsalizing activity in *Xenopus* embryos. We now designate the novel protein as "frzb-1." The gene for frzb-1 is expressed in many adult tissues of many animals, three of the cDNAs (*Xenopus*, mouse, and human) have been cloned by us. The accession numbers for the *Xenopus*, mouse, and human frzb-1 cDNA sequences of the gene now designated frzb-1 are U68059, U68058, and U68057, respectively. Frzb-1 has some degree of sequence similarity to the *Drosophila* gene frizzled which has been shown to encode a seven-transmembrane protein that can act both as a signalling and as a receptor protein (Vinson et al., *Nature*, 338, pp. 263-264, 1989; Vinson and Adler, *Nature*, 329, pp. 549-551, 1987). Vertebrate homologues of Frizzled have been isolated and they too were found to be anchored to the cell membrane by seven membrane spanning domains (Wang et al., *J. Biol. Chem.*, 271, pp. 4468-4476, 1996). Frzb-1 differs from the frizzled proteins in that it is an entirely soluble, diffusible secreted protein and therefore suitable as a therapeutic agent. The nucleotide sequence derived from *Xenopus* that, when

expressed, results in frzb-1 protein is illustrated by SEQ ID NO:4. The frzb-1 protein derived from mouse is shown as SEQ ID NO:7, while the mouse frzb-1 nucleotide sequence is SEQ ID NO:8. The human derived frzb-1 protein is illustrated by SEQ ID NO:9, and the human frzb-1 nucleotide sequence is SEQ ID NO:10.

Frzb-1 is an antagonist of Wnts *in vivo*, and thus is believed to find utility as a tumor suppressor gene, since over expressed Wnt proteins cause cancer. Frzb-1 may also be a useful vehicle for solubilization

and therapeutic delivery of Wnt proteins complexed with it.

The final cDNA isolated containing a signal sequence results in a peptide designated Paraxial
5 Protocadherin (PAPC). The cDNA for PAPC is a divergent member of the cadherin multigene family. PAPC is most related to protocadherin 43 reported by Sano et al., *The EMBO J.*, 12, pp. 2249-2256, 1993. As shown in SEQ ID NO:5, the PAPC gene encodes a transmembrane protein of
10 896 amino acids, of which 187 are part of an intracellular domain. PAPC is a cell adhesion molecule, and microinjection of PAPC mRNA constructs into *Xenopus* embryos suggest that PAPC acts as a molecule involved in mesoderm differentiation. A soluble form of the PAPC
15 extracellular domain is able to block muscle and mesoderm formation in *Xenopus* embryos. The nucleotide sequence encoding *Xenopus* PAPC is provided in SEQ ID NO:6.

Cerberus, frzb-1, or PAPC or fragments thereof
20 (which also may be synthesized by *in vitro* methods) may be fused (by recombinant expression or *in vitro* covalent methods) to an immunogenic polypeptide and this, in turn, may be used to immunize an animal in order to raise antibodies against the novel proteins. Antibodies
25 are recoverable from the serum of immunized animals. Alternatively, monoclonal antibodies may be prepared from cells from the immunized animal in conventional fashion. Immobilized antibodies are useful particularly in the diagnosis (*in vitro* or *in vivo*) or purification
30 of cerberus, frzb-1, or PAPC.

Substitutional, deletional, or insertional mutants of the novel polypeptides may be prepared by *in vitro* or recombinant methods and screened for immuno-crossreactivity with cerberus, frzb-1, or PAPC and for
35 cerberus antagonist or agonist activity.

Cerberus or frzb-1 also may be derivatized in vitro in order to prepare immobilized and labelled proteins, particularly for purposes of diagnosis of insufficiencies thereof, or for affinity purification of antibodies thereto.

Among applications for the novel proteins are tissue replacement therapy and, because frzb-1 is an antagonist of Wnt signaling, tumor suppression therapies. The cerberus receptor may define a novel signalling pathway. In addition, frzb-1 could permit the isolation of novel members of the Wnt family of growth factors.

Brief Description of the Drawings

Figure 1 illustrates the amino acid sequence (SEQ ID NO:1) of the Fig. 2 cDNA clone for cerberus;

Figure 2 illustrates a cDNA clone (SEQ ID NO:2) for cerberus derived from Xenopus. Sense strand is on top (5' to 3' direction) and the antisense strand on the bottom line (in the opposite direction);

Figures 3 and 4 show the amino acid and nucleotide sequence, respectively, of full-length frzb-1 from Xenopus (SEQ ID NOS:3 and 4);

Figures 5 and 6 show the amino acid and nucleotide sequence, respectively, of full-length PAPC from Xenopus (SEQ ID NOS:5 and 6);

Figures 7 and 8 show the amino acid and nucleotide sequence, respectively, of full-length frzb-1 from mouse (SEQ ID NOS:7 and 8); and

Figures 9 and 10 show the amino acid and nucleotide sequence, respectively, of full-length frzb-1 from human (SEQ ID NOS:9 and 10).

Detailed Description of the Preferred Embodiments

Among the several novel proteins and their nucleotide sequences described herein, is a novel endodermal, cardiac, and neural inducing factor in vertebrates that we have named "cerberus." When referring to cerberus, the present invention also contemplates the use of fragments, derivatives, agonists, or antagonists of cerberus molecules. Because cerberus has no homology to any reported growth factors, it is proposed to be the founding member of a novel family of growth factors with potent biological activities, which may be isolated using SEQ ID NO:2.

The amphibian organizer consists of several cell populations with region-specific inducing activities. On the basis of morphogenetic movements, three very different cell populations can be distinguished in the organizer. First, cells with crawling migration movements involute, fanning out to form the prechordal plate. Second, cells involute through the dorsal lip driven by convergence and extension movements, giving rise to the notochord of the trunk. Third, involution ceases and the continuation of mediolateral intercalation movements leads to posterior extension movements and to the formation of the tail notochord and of the chordoneural hinge. The three cell populations correspond to the head, trunk, and tail organizers, respectively.

The cerberus gene is expressed at the right time and place to participate in cell signalling by Spemann's organizer. Specifically, cerberus is expressed in the head organizing region that consists of crawling-migrating cells. The cerberus expressing region corresponds to the prospective foregut, including the liver and pancreas anlage, and the heart mesoderm.

Cerberus expression is activated by chordin, noggin, and organizer-specific homeobox genes.

Our studies were conducted in early embryos of the frog *Xenopus laevis*. The frog embryo is well suited to experiments, particularly experiments pertaining to generating and maintaining regional differences within the embryo for determining roles in tissue differentiation. It is easy to culture embryos with access to the embryos even at very early stages of development (preceding and during the formation of body pattern and differentiation) and the embryos are large. The initial work with noggin and chordin also had been in *Xenopus* embryos, and, as predicted, was highly conserved among vertebrates. Predictions based on work with *Xenopus* as to corresponding human noggin were proven true and the ability to clone the gene for human noggin was readily accomplished. (See the description of *Xenopus* work and cloning information in PCT application, published March 17, 1994, WO 9 405 800, and the subsequent human cloning based thereon in the PCT application, also published March 17, 1994, as WO 9 405 791.)

CLONING

The cloning of cerberus, frzb-1, and PAPC resulted from a comprehensive screen for cDNAs enriched in Spemann's organizer. Subtractive differential screening was performed as follows. In brief, poly A⁺ RNA was isolated from 300 dorsal lip and ventral marginal zone (VMZ) explants at stage 10½. After first strand cDNA synthesis approximately 70-80% of common sequences were removed by subtraction with biotinylated VMZ poly A⁺ RNA prepared from 1500 ventral gastrula halves. For differential screening, duplicate filters (2000 plaques per 15 cm plate, a total of 80,000 clones

screened) of an unamplified oriented dorsal lip library were hybridized with radiolabeled dorsal lip or VMZ cDNA. Putative organizer-specific clones were isolated, grouped by sequence analysis from the 5' end and whole-mount in situ hybridization, and subsequently classified into known and new dorsal-specific genes. Rescreening of the library (100,000 independent phages) with a cerberus probe resulted in the isolation of 45 additional clones, 31 of which had similar size as the longest one of the 11 original clones indicating that they were presumably full-length cDNAs. The longest cDNAs for cerberus, frzb-1, and PAPC were completely sequenced.

To explore the molecular complexity of Spemann's organizer we performed a comprehensive differential screen for dorsal-specific cDNAs. The method was designed to identify abundant cDNAs without bias as to their function. As shown in Table 1, five previously known cDNAs and five new ones were isolated, of which three (expressed as cerberus, frzb-1, and PAPC, respectively) had secretory signal sequences.

TABLE 1

	Previously Known Genes	Gene Product	No. of Isolates
	Chordin	novel secreted protein	70
	Goosecoid	homeobox gene	3
5	Pintallavis/XFKH-1	forkhead/transcription factor	2
	Xnot-2	homeobox gene	1
	Xlim-1	homeobox gene	1
	New Genes		
	Cerberus	novel secreted protein	11
10	PAPC	cadherin-like/transmembrane	2
	Frzb-1	novel secreted protein	1
	Sox-2	sry/transcription factor	1
	Fkh-like	forkhead/transcription factor	1

15 The most abundant dorsal-specific cDNA was chordin (chd), with 70 independent isolates. The second most abundant cDNA was isolated 11 times and named cerberus (after a mythological guardian dog with multiple heads). The cerberus cDNA encodes a putative secreted polypeptide of 270 amino acids, with an amino

20 terminal hydrophobic signal sequence and a carboxy terminal cysteine-rich region (Fig. 1). Cerberus is expressed specifically in the head organizer region of the *Xenopus* embryo, including the future foregut.

25 An abundant mRNA found in the dorsal region of the *Xenopus* gastrula encodes the novel putative secreted protein we have designated as cerberus. Cerberus mRNA has potent inducing activity in *Xenopus* embryos, leading to the formation of ectopic heads. Unlike other organizer-specific factors, cerberus does not dorsalize

30 mesoderm and is instead an inhibitor of trunk-tail mesoderm. Cerberus is expressed in the anterior-most

domain of the gastrula including the leading edge of the deep layer of the dorsal lip a region that, as shown here, gives rise to foregut and midgut endoderm. Cerberus promotes the formation of cement gland, olfactory placodes, cyclopic eyes, forebrain, and duplicated heart and liver (a foregut derivative). Because the pancreas is also derived from this foregut region, it is likely that cerberus induces pancreas in addition to liver. The expression pattern and inducing activities of cerberus suggest a role for a previously neglected region of the embryo, the prospective foregut endoderm, in the induction of the anterior head region of the embryo.

Turning to Fig. 1, *Xenopus cerberus* encodes a putative secreted protein transiently expressed during embryogenesis and the deduced amino acid sequence of *Xenopus cerberus* is shown. The signal peptide sequence and the nine cysteine residues in the carboxy-terminus are indicated in bold. Potential N-linked glycosylation sites are underlined. In database searches the cerberus protein showed limited similarity only to the mammalian Dan protein, a possible tumor suppressor proposed to be a DNA-binding protein.

Cerberus appears to be a pioneer protein, as its amino acid sequence and the spacing of its 9 cysteine residues were not significantly similar to other proteins in the databases (NCBI-Gen Bank release 93.0). We conclude that the second most abundant dorsal-specific cDNA encodes a novel putative secreted factor, which should be the founding member of a novel family of growth factors active in cell differentiation.

Cerberus Demarcates an Anterior Organizer Domain. Cerberus mRNA is expressed at low levels in the unfertilized egg, and zygotic transcripts start accumulating at early gastrula. Expression continues

during gastrula and early neurula, rapidly declining during neurulation. Importantly, cerberus expression starts about one hour after that of chd, suggesting that cerberus could act downstream of the chd signal.

5 Whole-mount *in situ* hybridizations reveal that expression starts in the yolky endomesodermal cells located in the deep layer of the organizer. The cerberus domain includes the leading edge of the most anterior organizer cells and extends into the lateral
10 mesoderm. The leading edge gives rise to liver, pancreas, and foregut in its midline, and the more lateral region gives rise to heart mesoderm at later stages of development.

Fig. 2 sets out the sequence of a full length
15 Xenopus cDNA for cerberus.

This entirely new molecule has demonstrated physiological properties that should prove useful in therapeutic, diagnostic, and clinical applications that require regeneration, differentiation, or repair of
20 tissues, such wound repair, neuronal regeneration or transplantation, supplementation of heart muscle differentiation, differentiation of pancreas and liver, and other applications in which cell differentiation processes are to be induced.

25 The second, novel, secreted protein we have discovered is called "frzb-1," which was shown to be a secreted protein in Xenopus oocyte microinjection experiments. Thus it provides a natural soluble form of the related extracellular domains of Drosophila and
30 vertebrate frizzled proteins. We propose that the latter proteins could be converted into active soluble forms by introducing a stop codon before the first transmembrane domain. We have noted that the cysteine-rich region of frzb-1 and frizzl d contains some overall
35 structural homology with Wnt proteins using the Profile

Search homology program (Gribskov, *Meth. Enzymol.*, 183, pp. 146-159, 1990). This had raised the interesting possibility that frzb-1 could interact directly with Wnt growth factors in the extracellular space. This was
5 because we had found that when microinjected into *Xenopus* embryos, frzb-1 constructs have moderate dorsalizing activity, leading to the formation of embryos with enlarged brain and head, and shortened trunk. Somatic muscle differentiation, which requires
10 Xwnt-8, was inhibited. In the case of frzb-1, an attractive hypothesis, suggested by the structural homologies, was that it may act as an inhibitor of Wnt-8, a growth factor that has ventralizing activity in the *Xenopus* embryo (Christian and Moon, *Genes Dev.*, 7,
15 pp. 13-28, 1993). We have shown that frzb-1 can interact with Xwnt-8 and Wnt-1, and it is expected that it could also interact with other members of the Wnt family of growth factors, of which at least 15 members exist in mammals. In addition, a possible interaction
20 with Wnts was suggested by the recent discovery that dishevelled, a gene acting downstream of wingless, has strong genetic interaction with frizzled mutants in *Drosophila* (Krasnow et al., *Development*, 121, pp. 4095-4102, 1995). This possibility has been explored in
25 depth (Leyns et al., *Cell*, 88, pp. 747-756, March 21, 1997), because a soluble antagonist of the Wnt family of proteins is expected to be of great therapeutic value. Examples 1 and 2 illustrate tests that show antagonism of Xwnt-8 by binding to frzb-1.

30 Vertebrate homologues of Frizzled have been isolated and they too are anchored to the cell membrane by seven membrane spanning domains (Wang et al., *J. Biol. Chem.*, 271, pp. 4468-4476, 1996). Frzb-1 differs from the frizzled proteins in that it is an
35 entirely soluble, diffusible secreted protein and

therefore suitable as a therapeutic agent. The nucleotide sequence that when expressed results in frzb-1 protein is illustrated by SEQ ID NO:4.

SEQ ID NO:4 corresponds to the *Xenopus* homolog, but by using it in BLAST searches (and by cloning mouse frzb-1) we had been able to assemble the sequence of the entire mature human frzb-1 protein, SEQ ID NO:9. Indeed, human frzb-1 is encoded in six expressed sequence tags (ESTs) available in Genbank.

10 The human frzb-1 sequence can be assembled by overlapping in the 5' to 3' direction the ESTs with the following accession numbers in Genbank: H18848, R63748, W38677, W44760, H38379, and N71244. No function had yet been assigned to these EST sequences, but we

15 believe and thus propose here that human frzb-1 will have similar functions in cell differentiation to those described above for *Xenopus* frzb-1. The nucleotide sequence of human frzb-1 is shown in SEQ ID NO:10. The mouse frzb-1 protein and nucleotide sequences are

20 provided by SEQ ID NOS:7 and 8, respectively.

In particular, we believe that frzb-1 will prove useful in gene therapy of human cancer cells. In this rapidly developing field, one approach is to introduce vectors expressing anti-sense sequences to

25 block expression of dominant oncogenes and growth factor receptors. Another approach is to produce episomal vectors that will replicate in human cells in a controlled fashion without transforming the cells. For an example of the latter (an episomal expression vector

30 system for human gene therapy), reference is made to U.S. Patent 5,624,820, issued April 29, 1997, inventor Cooper.

Gene therapy now includes uses of human tumor suppression genes. For example, U.S. Patent 5,491,064,

35 issued February 13, 1996, discloses a tumor suppression

gene localized on chromosome 11 and described as potentially useful for gene therapy in cancers deleted or altered in their expression of that gene. Frzb-1 maps to chromosome 2q31-33 and loss of one copy of the
5 2q31-33 and loss of one copy of the 2q arm has been observed with high incidence in lung carcinomas, colo-rectal carcinomas, and neuroblastomas, which has lead to the proposal that the 2q arm carries a tumor suppressor gene. We expect frzb to be a tumor
10 suppressor gene, and thus to be useful in tumor suppression applications.

A number of applications for cerberus and frzb-1 are suggested from their pharmacological (biological activity) properties.

15 For example, the cerberus and frzb-1 cDNAs should be useful as a diagnostic tool (such as through use of antibodies in assays for proteins in cell lines or use of oligonucleotides as primers in a PCR test to amplify those with sequence similarities to the
20 oligonucleotide primer, and to determine how much of the novel protein is present).

Cerberus, of course, might act upon its target cells via its own receptor. Cerberus, therefore, provides the key to isolate this receptor. Since many

25 receptors mutate to cellular oncogenes, the cerberus receptor should prove useful as a diagnostic probe for certain tumor types. Thus, when one views cerberus as ligand in complexes, then complexes in accordance with the invention include antibody bound to cerberus,
30 antibody bound to peptides derived from cerberus, cerberus bound to its receptor, or peptides derived from cerberus bound to its receptor or other factors. Mutant forms of cerberus, which are either more potent agonists or antagonists, are believed to be clinically useful.

Such complexes of cerberus and its binding protein partners will find uses in a number of applications.

Practice of this invention includes use of an oligonucleotide construct comprising a sequence coding
5 for cerberus or frzb-1 and for a promoter sequence
operatively linked in a mammalian or a viral expression
vector. Expression and cloning vectors contain a
nucleotide sequence that enables the vector to replicate
in one or more selected host cells. Generally, in
10 cloning vectors this sequence is one that enables the
vector to replicate independently of the host
chromosomes, and includes origins of replication or
autonomously replicating sequences. The well-known
plasmid pBR322 is suitable for most gram negative
15 bacteria, the 2 μ plasmid origin for yeast and various
viral origins (SV40, polyoma, adenovirus, VSV or BPV)
are useful for cloning vectors in mammalian cells.

Expression and cloning vectors should contain
a selection gene, also termed a selectable marker.
20 Typically, this is a gene that encodes a protein
necessary for the survival or growth of a host cell
transformed with the vector. The presence of this gene
ensures that any host cell which deletes the vector will
not obtain an advantage in growth or reproduction over

25 transformed hosts. Typical selection genes encode
proteins that (a) confer resistance to antibiotics or
other toxins, e.g. ampicillin, neomycin, methotrexate or
tetracycline, (b) complement auxotrophic deficiencies.

Examples of suitable selectable markers for
30 mammalian cells are dihydrofolate reductase (DHFR) or
thymidine kinase. Such markers enable the identifica-
tion of cells which were competent to take up the
cerberus nucleic acid. The mammalian cell transformants
ar placed under selection pressure which only the
35 transf rmants are uniquely adapted to survive by virtue

of having taken up the marker. Selection pressure is imposed by culturing the transformants under conditions in which the concentration of selection agent in the medium is successively changed. Amplification is the process by which genes in greater demand for the production of a protein critical for growth are reiterated in tandem within the chromosomes of successive generations of recombinant cells. Increased quantities of cerberus or frzb-1 can therefor be synthesized from the amplified DNA.

For example, cells transformed with the DHFR selection gene are first identified by culturing all of the transformants in a culture medium which contains methotrexate (Mtx), a competitive antagonist of DHFR. An appropriate host cell in this case is the Chinese hamster ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub and Chasin, *Proc. Nat. Acad. Sci.*, 77, 4216 (1980). The transformed cells then are exposed to increased levels of Mtx. This leads to the synthesis of multiple copies of the DHFR gene and, concomitantly, multiple copies of other DNA comprising the expression vectors, such as the DNA encoding cerberus or frzb-1. Alternatively, host cells transformed by an expression vector comprising DNA sequences encoding cerberus or frzb-1 and aminoglycoside 3' phosphotransferase (APH) protein can be selected by cell growth in medium containing an aminoglycosidic antibiotic such as kanamycin or neomycin or G418. Because eukaryotic cells do not normally express an endogenous APH activity, genes encoding APH protein, commonly referred to as neo resistant genes, may be used as dominant selectable markers in a wide range of eukaryotic host cells, by which cells transformed by the vector can readily be identified.

Expression vectors, unlike cloning vectors, should contain a promoter which is recognized by the host organism and is operably linked to the cerberus nucleic acid. Promoters are untranslated sequences located upstream from the start codon of a structural gene (generally within about 100 to 1000 bp) that control the transcription and translation of nucleic acid under their control. They typically fall into two classes, inducible and constitutive. Inducible promoters are promoters that initiate increased levels of transcription from DNA under their control in response to some change in culture conditions, e.g. the presence or absence of a nutrient or a change in temperature. At this time a large number of promoters recognized by a variety of potential host cells are well known. These promoters can be operably linked to cerberus encoding DNA by removing them from their gene of origin by restriction enzyme digestion, followed by insertion 5' to the start codon for cerberus or frzb-1.

Nucleic acid is operably linked when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein which participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, operably linked means that the DNA sequences being linked are contiguous and, in the case of a secretory leader, contiguous and in reading phase. Linking is accomplished by ligation at convenient restriction sites. If such sites do not

exit then synthetic oligonucleotide adapters or linkers are used in accord with conventional practice.

Transcription of the protein-encoding DNA in mammalian host cells is controlled by promoters obtained
5 from the genomes of viruses such as polyoma, cytomegalovirus, adenovirus, retroviruses, hepatitis-B virus, and most preferably Simian Virus 40 (SV40), or from heterologous mammalian promoters, e.g. the actin promoter. Of course, promoters from the host cell or
10 related species also are useful herein.

Cerberus and frzb-1 are clearly useful as a component of culture media for use in culturing cells, such as endodermal, cardiac, and nerve cells, *in vitro*. We believe cerberus and frzb-1 will find uses as agents
15 for enhancing the survival or inducing the growth of liver, pancreas, heart, and nerve cells, such as in tissue replacement therapy.

The final cDNA isolated containing a signal sequence results in a peptide designated Paraxial Protocadherin (PAPC). The cDNA for PAPC is a divergent member of the cadherin multigene family. PAPC is most related to protocadherin 43 reported by Sano et al., *The EMBO J.*, 12, pp. 2249-2256, 1993. As shown in SEQ ID NO:5, the PAPC gene encodes a transmembrane protein of

25 896 amino acids, of which 187 are part of an intracellular domain. PAPC is a cell adhesion molecule, and microinjection of PAPC mRNA constructs into *Xenopus* embryos suggest that PAPC acts in mesoderm differentiation. The nucleotide sequence encoding
30 *Xenopus* PAPC is provided in SEQ ID NO:6.

Therapeutic formulations of the novel proteins may be prepared for storage by mixing the polypeptides having the desired degree of purity with optional physiologically acceptable carriers, excipients or
35 stabilizers, in the form of lyophilized cake or aqueous

solutions. Acceptable carriers, excipients or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; anti-oxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin or immunoglobulins. Other components can include glycine, glutamine, asparagine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as Tween, Pluronic or PEG.

Polyclonal antibodies to the novel proteins generally are raised in animals by multiple subcutaneous (sc) or intraperitoneal (ip) injections of cerberus or frzb-1 and an adjuvant. It may be useful to conjugate these proteins or a fragment containing the target amino acid sequence to a protein which is immunogenic in the species to be immunized, e.g., keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, or soybean trypsin inhibitor using a bifunctional or derivatizing agent, for example, maleimidobenzoyl sulfosuccinimide ester (conjugation through cysteine residues), N-hydroxysuccinimide (through lysine residues), glutaraldehyde, succinic anhydride, SOCl_2 , or $\text{R}^1\text{N} = \text{C} = \text{NR}$.

Animals can be immunized against the immunogenic conjugates or derivatives by combining 1 mg or 1 μg of conjugate (for rabbits or mice, respectively) with 3 volumes of Freund's complete adjuvant and injecting the solution intradermally in multiple sites. One month later the animals are boosted with 1/5 to 1/10 the original amount of conjugate in Freund's complete

adjuvant by subcutaneous injection at multiple sites. Seven to 14 days later animals are bled and the serum is assayed for anti-cerberus titer. Animals are boosted until the titer plateaus. Preferably, the animal is
5 boosted with the conjugate of the same cerberus or frzb-1 polypeptide, but conjugated to a different protein and/or through a different cross-linking agent. Conjugates also can be made in recombinant cell culture as protein fusions. Also, aggregating agents such as
10 alum are used to enhance the immune response.

Monoclonal antibodies are prepared by recovering spleen cells from immunized animals and immortalizing the cells in conventional fashion, e.g. by fusion with myeloma cells or by EB virus transformation
15 and screening for clones expressing the desired antibody.

Antibodies are useful in diagnostic assays for cerberus, frzb-1, or PAPC or their antibodies and to identify family members. In one embodiment of a
20 receptor binding assay, an antibody composition which binds to all of a selected plurality of members of the cerberus family is immobilized on an insoluble matrix, the test sample is contacted with the immobilized antibody composition in order to adsorb all cerberus

25 family members, and then the immobilized family members are contacted with a plurality of antibodies specific for each member, each of the antibodies being individually identifiable as specific for a predetermined family member, as by unique labels such as
30 discrete fluorophores or the like. By determining the presence and/or amount of each unique label, the relative proportion and amount of each family member can be determined.

The antibodies also are useful for the
35 affinity purification of the novel proteins from

recombinant cell culture or natural sources. Antibodies that do not detectably cross-react with other growth factors can be used to purify the proteins free from these other family members.

5

EXAMPLE 1

Frzb-1 Antagonizes Xwnt-8 Non-Cell Autonomously

To test whether frzb-1 can antagonize secondary axes caused by Xwnt-8 after secretion by injected cells, an experimental design was used. Thus, frzb-1 mRNA was injected into each of the four animal blastomeres of eight-cell embryos, and subsequently, a single injection of Xwnt-8 mRNA was given to a vegetal-ventral blastomere at the 16-32 cell stage. In two independent experiments, we found that injection of frzb-1 alone (n=13) caused mild dorsalization with enlargement of the cement gland in all embryos and that injection of Xwnt-8 alone (n=53) lead to induction of complete secondary axes in 67% of the embryos. However, injection of frzb-1 into animal caps abolished the formation of complete axes induced by Xwnt-8 (n=27), leaving only a residual 14% of embryos with very weak secondary axes. The double-injected embryos retained the enlarged cement gland phenotype caused by injection of frzb-1 mRNA alone. Because both mRNAs encode secreted proteins and were microinjected into different cells, we conclude that the antagonistic effects of frzb-1 and Xwnt-8 took place in the extracellular space after these proteins were secreted.

EXAMPLE 2

Membrane-Anchored Wnt-1 Confers Frzb-1 Binding

To investigate a possible interaction between frzb-1 and Wnts, the first step was to insert an HA epitope tag into a Xenopus frzb-1 construct driven by the CMV (cytomegalovirus) promoter. Frzb1-HA was tested in mRNA microinjection assays in Xenopus embryos and found to be biologically active. Conditioned medium from transiently transfected cells contained up to 10 μ g/ml of Frzb1-HA (quantitated on Western blots using an HA-tagged protein standard).

Transient transfection of 293 cells has been instrumental in demonstrating interactions between wingless and frizzled proteins. We therefore took advantage of constructs in which Wnt-1 was fused at the amino terminus of CD8, generating a transmembrane protein containing biologically active Wnt-1 exposed to the extracellular compartment. A Wnt1CD8 cDNA construct (a generous gift of Dr. H. Varmus, NIH) was subcloned into the pCDNA (Invitrogen) vector and transfected into 293 cells. After incubation with Frzb1-HA-conditioned medium (overnight at 37°C), intensely labeled cells were observed by immunofluorescence. As a negative control, a construct containing 120 amino acids of Xenopus chordin, an unrelated secreted protein was used. Transfection of this construct produced background binding of Frzb1-HA to the extracellular matrix, both uniform and punctate. Cotransfection of Wnt1CD8 with pCDNA-LacZ showed that transfected cells stained positively for Frzb1-HA and LacZ. Since Wnt1CD8 contains the entire CD8 molecule, a CD8 cDNA was used as an additional negative control. After transfection with LacZ and full-length CE8, Frzb1-HA failed to bind to the transfected cells. Although most of our experiments

were carried out at 37°C, Frzb1-HA-conditioned medium also stained Wnt1CD8-transfected cells after incubation at 4°C for 2 hours.

Attempts to biochemically quantitate the binding of Frzb-1 to Wnt1CD8-transfected cells were unsuccessful due to high background binding to control cultures, presumably due to binding to the extracellular matrix. Thus, we were unable to estimate a K_D for the affinity of the Frzb-1/Wnt-1 interaction. However, when serial dilutions of conditioned medium containing Frzb1-HA were performed (ranging from 2.5×10^{-7} to 1.25×10^{-10} M), staining of Wnt1CD8-transfected cells was found at all concentrations.

Although we have been unable to provide biochemical evidence for direct binding between Wnts and frzb-1, this cell biological assay indicates that Frzb1-HA can bind, directly or indirectly, to Wnt-1 on the cell membrane in the 10^{-10} M range.

It is to be understood that while the invention has been described above in conjunction with preferred specific embodiments, the description and examples are intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims.

It is Claimed:

1. A substantially pure protein characterized by a physiologically active form and comprising an amino acid sequence encoded by the DNA of SEQ ID NO:2.

2. The protein as in claim 1 having neurotrophic, growth or differentiation factor activity.

3. A composition comprising the protein of claim 1 and a physiologically acceptable carrier with which the peptide is admixed.

4. An oligonucleotide construct comprising a sequence coding for a protein and an expression vector operatively linked therewith, the protein having neurotrophic, growth or differentiation factor activity
5 and being expressible from SEQ ID NO:2.

5. The construct as in claim 4 wherein the expression vector is a mammalian or viral expression vector.

6. A substantially pure protein characterized by a physiologically active form and comprising an amino acid sequence encoded by the DNA of SEQ ID NO:4, SEQ ID NO:8, or SEQ ID NO:10.

7. The protein as in claim 6 having neurotrophic, growth or differentiation factor activity.

8. A composition comprising the protein of claim 6 and a physiologically acceptable carrier with which the protein is admixed.

9. An oligonucleotide construct comprising a sequence coding for a protein and an expression vector operatively linked therewith, the protein being expressible from SEQ ID NO:4, SEQ ID NO:8 or SEQ ID NO:10.

10. The construct as in claim 9 wherein the protein is expressible in soluble form.

11. The construct as in claim 9 wherein the expression vector is a mammalian or viral expression vector.

12. A complex comprising a substantially pure frzb-1 protein complexed with at least one Wnt protein.

13. A substantially pure protein characterized by a physiologically active form and comprising an amino acid sequence encoded by the DNA of SEQ ID NO:6.

14. The protein as in claim 13 having mesoderm differentiation activity.

15. A composition comprising the protein of claim 13 and a physiologically acceptable carrier with which the protein is admixed.

MLLNVLRI	CI	IVCLVNDGAG	KHSEGRERTK	TYSLNSRGYF	40
RKERGARRSK	ILLVNTKGLD	EPHIGHGDFG	LVAELFDSTR	80	
THTNRKEPDM	NKVLFSTVA	HGNKSARRKA	YNGSRRNIFS	120	
RRSFDKRNTE	VTEKPGAKMF	WNNFLVKMNG	APQNTSHGSK	160	
AQEIMKEACK	TLPFTQNI	VH	ENCDRMVION	NLCFGK CISL	200
HVPNQD	RRN	TCSHCLPSKF	TLNHLTLNCT	GSKNVVKVVM	240
MVEECTCEAH	KSNFHQTAQF	NMDTSTTLHH			270

Figure 1

SUBSTITUTE SHEET (RULE 26)

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GAATTCCAG CAAGTCGCTC AGAAACACTG CAGGGTCTAG ATATCATACA ATGTTACTAA CTTAAGGGTC GTTCAGCGAG TCTTTGTGAC GTCCAGATC TATAGTATGT TACAATGATT	60
ATGTAICTAG GATCTGTATT ATCGTCTGCC TTGTGAATGA TGGAGCAGGA AAACACTCAG TACATGAGTC CTAGACATAA TAGCAGACGG AACACTTACT ACCTCGTCCT TTTGTGAGTC	120
AAGGACGAGA AAGGACAAAA ACATATTAC TTAACAGCAG AGGTTACTTC AGAAAAGAA TTCCTGCTCT TTCCTGTTTT TGTATAAGTG AATTGTGCTC TCCAATGAAG TCTTTTCTTT	180
GAGGAGCAG TAGGAGCAAG ATTCTGCTGG TGAATACTAA AGGTCTTGAT GAACCCACA CTCCTGTCG ATCCTGCTTC TAAGACGACC ACTTATGATT TCCAGAATA CTGGGGTGT	240
TTGGGCATGG TGATTTTCGC TTAGTAGCTG AACTATTTGA TTCCACCAGA ACACATACAA AACCCGTACC ACTAAAAGCG AATCATGAC TTGATAAACT AAGGTGGTCT TGTGTATGTT	300
ACAGAAAAGA GCCAGACATG AACAAAGTCA AGCTTTTCTC AACAGTTGCC CATGGAAACA TGTCTTTCT CGGTCTGTAC TTGTTTCAGT TCGAAAAGAG TTGTCAACGG GTACCTTTGT	360
AAAGTGCAAG AAGAAAAGCT TACAATGGTT CTAGAAGGAA TATTTTCTCT CGCCGTCTCT TTTACGTTT TTTCTTTTGA ATGTTACCA GATCTTCTT ATAAAAAGGA GCGGCAAGAA	420
TTGATAAAG AAATACAGAG GTTACTGAAA AGCCTGGTGC CAAGATGTTT TGGAACAATT AACTATTTT TTTATGTCTC CAATGACTTT TCGGACCAAG GTTCTACAAG ACCTTGTTAA	480
TTTTGGTTAA AATGAATGGA GCCCCACAGA ATACAAGCCA TGGCAGTAAA GCACAGGAAA AAAACCAATT TTACTTACCT CGGGGTGTCT TATGTTGGT ACCGTCATTT CGTGTCTTTT	540
TAATGAAAGA AGCTTGCAA ACCTTGTTTT TCACTCAGAA TATTGTACAT GAAAACGTG ATTACTTTCT TCGAACGTTT TGGAAACAAA AGTGAGTCTT ATACATGTA CTTTGTACAC	600
ACAGGATGGT GATACAGAAC AATCTGTGCT TTGGTAAATG CATCTCTCTC CATGTTCCAA TGTCCTACCA CTATGTCTTG TTAGACAAGA AACCATTTAC GTAGAGAGAG GTACAAGGTT	660
ATCAGCAAGA TCGACGAAAT ACTTGTTGCC ATTGCTTGCC GTCCAAATTT ACCCTGAACC TAGTGTCTCT AGCTGCTTTA TGAACAGGG TAACGAAACG CAGGTTTAAA TGGGACTTGG	720
ACCTGACGCT GAATTGTACT GGATCTAAGA ATGTAGTAAA GGTGTGATG ATGGTAGAGG TGGACTGCGA CTTAACATGA OCTAGATTCT TACATCATT CCAACAGTAC TACCATCTCC	780
AATGCACTG TGAAGCTCAT AAGAGCAACT TCACCAAAAC TGCACAGTTT AACATGGATA TTACGTGCAC ACTTCGAGTA TTCTCGTTGA AGGTGGTTTG ACGTGTCAAA TTGTACCTAT	840
CATCTACTAC CCTGCACCAT TAAAGGACTG CCATACAGTA TGGAAATGCC CTTTTGTTGG GTAGATGATG GGACGTGGTA ATTTCTGAC GGTATGTCAT ACCTTTACGG GAAAACAACC	900
AATATTTGTT ACATACTATG CATCTAAAGC ATTATGTTGC CTTCTATTTT ATATAACCAC TTATAAACAA TGTATGATAC GTAGATTTTC TAATACAACG GAAGATAAAG TATATTGGTG	960
ATGGAATAAG GATTGTATGA ATTATAATTA ACAAATGGCA TTTTGTGTAA CATGCAAGAT TACCTTATTC CTAACATACT TAATATTAAT TGTTTACCGT AAAACACATT GTACGTTCTA	1020

Figure 2A

SUBSTITUTE SHEET (RULE 28)

CTCTGTTCCA TCAGTTGCAA GATAAAAGGC AATATTTGTT TGACTTTTTT TCTACAAAT 1080
GAGACAAGGT AGTCAACGTT CTATTTTCCG TTATAAACAA ACTGAAAAAA AGATGTTTTA

GAATACCCAA ATATATGATA AGATAATGGG GTCAAACTG TTAAGGGGTA ATGTAATAAT 1140
CTTATGGGTT TATATACTAT TCTATTACCC CAGTTTTGAC AATTCCCAT TACATTATTA

AGGGACTAAG TTTGCCAGG AGCAGTGACC CATAACAACC AATCAGCAGG TATGATTTAC 1200
TCCCTGATTC AAACGGGTCC TCGTCACTGG GTATTGTTGG TTAGTGTGCC ATACTAAATG

TGGTCACCTG TTTAAAAGCA AACATCTTAT TGGTTGCTAT GGGTTACTGC TTCTGGGCAA 1260
ACCAGTGGAC AAATTTTCGT TTGTAGAATA ACCAACGATA CCCAATGACG AAGACCCGTT

AATGTGTGCC TCATAGGGGG GTTAGTGTGT TGTGTACTGA ATAAATTGTA TTTATTTTCAT 1320
TTACACACGG AGTATCCCC CAATCACACA ACACATGACT TATTTAACAT AAATAAAGTA

TGTTACAAA AAAAAAA
ACAATGTTTT TTTTTTT

Figure 2B

SUBSTITUTE SHEET (RULE 26)

MSRTRKVDL LLLAIPGLAL LLLPNAYCAS CEPVRIPMCK SMPWNMTKMP NHLHHSTQAN	60
AILAIEQFEG LLTTECSQDL LFFLCAMYAP ICTIDFQHEP IKPCKSV CER ARAGCEPILI	120
KYRHTWPESL ACEELPVYDR GVCISPEAIV TVEQGTDSMP DFSMDSNNGN CGSGREHCKC	180
KPMKATQKTY LKNNYNYVIR AKVKEVKVVC HDATAIVEVK EILKSSLVNI PKDTVTLYTN	240
SGCLCPQLVA NEEYIIMGYE DKERTRLLLV EGSLAEKWRD RLAKVKRWD QKLRRPRKSK	300
DPVAPIPNKN SNSRQARS	

Figure 3

SUBSTITUTE SHEET (RULE 26)

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GAATTCOCTT TCACACAGGA CTCTGGCAG AGGTGAATGG TTAGCCCTAT GGATTTGGTT	60
CTTAAGGGAA AGTGTGTCTT GAGGACCGTC TCCACTTACC AATCGGGATA CCTAAACCAA	
TGTTGATTTT GACACATGAT TGATTGCTTT CAGATAGGAT TGAAGGACTT GGATTTTAT	120
ACAACTAAAA CTGTGTACTA ACTAACGAAA GTCTATCCTA ACTTCCTGAA CCTAAAAATA	
CTAATTCTGC ACTTTTAAAT TATCTGAGTA ATTGTTCAAT TTGTATTGGA TGGGACTAAA	180
GATTAAGACG TGAAAATTTA ATAGACTCAT TAACAAGTAA AACATAACCT ACCCTGATTT	
GATAAACTTA ACTCCTTGCT TTTGACTTGC CCATAAACTA TAAGGTGGGG TGAGTTGTAG	240
CTATTTGAAT TGAGGAACGA AACTGGAACG GGTATTTGAT ATTCCACCCC ACTCAACATC	
TTGCTTTTAC ATGTGCCCAG ATTTTCCTCG TATTCCCTGT ATTCCCTCTA AAGTAAGCCT	300
AACGAAAATG TACACGGGTC TAAAAGGGAC ATAAGGGACA TAAGGGAGAT TTCATTCCGA	
ACACATACAG GTTGGGCAGA ATAACAATGT CTGGAACAAG GAAAGTGGAC TCATTACTGC	360
TGTGTATGTC CAACCCGTCT TATTGTTACA GAGCTTGTTT CTTTCACTG AGTAATGACG	
TACTGGOCAT ACCTGGACTG GCGCTTCTCT TATTACCCAA TGCTTACTGT GCTTCGTGTG	420
ATGACCGGTA TGAOCTGAC CGGAAGAGA ATAATGGGT ACGAATGACA CGAAGCACAC	
AGCCTGTGCG GATCCCCATG TGCAATCTA TGCCATGGAA CATGACCAAG ATGCCCAACC	480
TGGACACGC CTAGGGGTAC ACGTTTAGAT ACGGTACCTT GTACTGGTTC TACGGGTTGG	
ATCTCCACCA CAGCACTCAA GCCAATGCCA TCCTGGCAAT TGAACAGTTT GAAGGTTTGC	540
TAGAGGTGGT GTCGTGAGTT CCGTTACGGT AGGACCGTTA ACTTGTCAA CTTCCAAACG	
TGACCACTGA ATGTAGCCAG GACCTTTTGT TCTTTCTGTG TGCCATGTAT GCCCCATT	600
ACTGGTGACT TACATCGGTC CTGGAAAACA AGAAGACAC ACGGTACATA CGGGGTAAA	
GTACCATCGA TTTCAGCAT GAACCAATTA AGCCTTGCAA GTCCGTGTGC GAAAGGGCCA	660
CATGGTAGCT AAAGGTGTA CTTGGTTAAT TCGGAACGTT CAGGCACACG CTTTCCCGGT	
GGCCCGGCTG TGAGCCCAT CTCTATAAGT ACCGGCACAC TTGGCCAGAG AGCCTGGCAT	720
CCGGCCGAC ACTCGGTAA GAGTATTCA TGGCCGTGTG AACCGGTCTC TCGGACCGTA	
GTGAAGAGCT GCGGTATAT GACAGAGGAG TCTGCATCTC CCGAGAGGCT ATCGTCACAG	780
GACTTCTGGA GCGGCATATA CTGTGTGCTG AGAGGTGAG GGGTCTGGA TAGCACTGTC	
TGGAACAAGG AACAGATTCA ATGCCAGACT TCTCATGGA TTCAAACAAT GGAAATTGCG	840
ACCTTGTTCC TTGTCTAAGT TACGGTCTGA AGAGGTACCT AAGTTTGTTA CCTTTAAGC	
GAAGGGGCGG GGAGCACTGT AAATGCAAGC CCATGAAGGC AACCCAAAAG ACGTATCTCA	900
CTTCGCGGTC CCTCGTGACA TTTACGTTTG GGTACTTCCG TTGGGTTTTT TGCATAGAGT	
AGAATAATTA CAATTATGTA ATCAGAGCAA AAGTGAAAGA GGTGAAAGTG AAATGCCACG	960
TCTTAITTAAT GTTAATACAT TAGTCTGTTT TTCATTTTCT CCACTTTTAC TTTACGGTGC	
ACGCAACAGC AATTGTGGAA GTAAAGGAGA TTCTCAAGTC TTCCCTAGTG AACATTCTTA	1020
TGCGTTGTG TTAACACCTT CATTCCTCT AAGAGTTCAG AAGGGATCAC TTGTAAGGAT	

Figure 4A

SUBSTITUTE SHEET (RULE 26)

AAGACACAGT GACACTGTAC ACCAACTCAG GCTGCTTGTG CCCCCAGCTT GTTGCCAATG 1080
 TTCTGTGTCA CTGTGACATG TGGTTGAGTC CGACGAACAC GGGGGTGGAA CAACGGTTAC
 AGGAATACAT AATTATGGGC TATGAAGACA AAGAGCGTAC CAGGCTTCTA CTAGTGGAAG 1140
 TCCTTATGTA TTAATACCG ATACTTCTGT TTCTCGCATG GTCOGAAGAT GATCACCTTC
 GATCCTTGGC CGAAAAATGG AGAGATCGTC TTGCTAAGAA AGTCAAGCGC TGGGATCAAA 1200
 CTAGGAACCG GCTTTTTACC TCTCTAGCAG AACGATTCTT TCAGTTCGCG ACCCTAGTTT
 AGCTTCGACG TCCCAGGAAA AGCAAAGACC CCGTGGCTCC AATTCCCAAC AAAACAGCA 1260
 TCGAAGCTGC AGGGTCCTTT TCGTTTCTGG GGCACCGAGG TTAAGGGTTG TTTTGTCTGT
 ATTCCAGACA AGCGCGTAGT TAGACTAAGC GAAAGGTGTA TGGAACTCT ATGGACTTTG 1320
 TAAGGTCTGT TCGGCGATCA ATCTGATTGC CTTTCCACAT ACCTTTGAGA TACCTGAAAC
 AACTAAGAT TTGCATTGTT GGAAGAGCAA AAAAGAAATT GCACTACAGC ACGTTATATT 1380
 TTTGATTCTA AACGTAACAA CCTTCTCGTT TTTTCTTAA CGTGATGTCTG TGCAATATAA
 CTATTGTTA CTACAAGAAG CTGGTTAGT TGATTGTAGT TCTCCTTCC TTCTTTTTT 1440
 GATAACAAAT GATGTTCTTC GACCAATCA ACTAATCATCA AGAGGAAAGG AAGAAAAAAA
 TTATAACTAT ATTTGCACGT GTTCCAGGC AATTGTTTTA TTCAACTTCC AGTGACAGAG 1500
 AATATTGATA TAAAGTGCA CAAGGGTCCG TTAACAAAT AAGTTGAAGG TCACTGTCTC
 CAGTGACTGA ATGTCTCAGC CTAAAGAAGC TCAATTCATT TCTGATCAAC TAATGGTGAC 1560
 GTCACTGACT TACAGAGTCG GATTCTTCTG AGTTAAGTAA AGACTAGTTG ATTACCACTG
 AAGTGTITGA TACTTGGGGA AAGTGAAC TAAGCAATGG TAAATCAGAG AAAAGTTGAC 1620
 TTCACAACT ATGAACCCCT TTCACTTGAT TAAGTTAOC ATTTAGTCTC TTTTCAACTG
 CAATGTTGCT TTTCTGTAG ATGAACAAGT GAGAGATCAC ATTTAAATGA TGATCACTTT 1680
 GTTACAAOGA AAAGGACATC TACTTGTTCA CTCTCTAGTG TAAATTTACT ACTAGTGAAA
 CCATTTAATA CTTTCAGCAG TTTTAGTTAG ATGACATGTA GGATGCACCT AAATCTAAAT 1740
 GGTAAATTAT GAAAGTCGTC AAAATCAATC TACTGTACAT CCTACGTGGA TTTAGATTTA
 ATTTATCAT AAATGAAGAG CTGGTTTGA CTGTATGGTC ACTGTTGGGA AGGTAAATGC 1800
 TAAATAGTA TTTACTTCTC GACCAATCT GACATACCAG TGACAACCT TCCATTTACG
 CTACTTTGTC AATTCTGTTT TAAAAATTGC CTAAATAAAT ATTAAGTCCT AAATAAAAAA 1860
 GATGAACAG TTAAGACAAA ATTTTAAAG GATTATTTA TAATTCAGGA TTTATTTTTT

AAAAAAAAAA AAAAA
 TTTTTTTTTT TTTT

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MLLLFRAIPM LLLGLMVLQT DCEIAQYYID EEEPPGTVIA VLSQHSIFNT TDIPATNFRL	60
MKQFNNSLIG VRESQGQLSI MERIDREQIC RQSLHCNLAL DVVSFSKGHF KLLNVKVEVR	120
DINDHSPHEP SEIMHVEVSE SSSVGTRIPL EIAIDEDVGS NSIQNFQISN NSHFSIDVLT	180
RADGVKYADL VLMRELDREI OPTYIMELLA MDGGVPSLSG TAVVNIRVLD FNDNSPVFER	240
STIAVDLVED APLGYLLEL HATDDDEGVN GEIVYGFSTL ASQEVRLFK INSRTGSVTL	300
EGQVDFETKQ TYEFEVQAQD LGPNPLTATC KVTVHILDVN DNTPAITITP LTTVNAGVAY	360
IPETATKENF IALISTTDRA SGSNGQVRCT LYGHEHFKLQ QAYEDSYMIV TTSTLDRENI	420
AAYSLTVVAE DLGFPSLTKK KYITVKVSDE NDNAPVFSKP QYEASILENN APGSYITTVI	480
ARDSSDQNG KVNRLVDAK VMGQSLTTFV SLDADSGVLR AVRSLDYEKL KQLDFEIEAA	540
DNGIPQLSTR VQNLRLIVDQ NDNCPVITNP LLNNGSGEVL LPISAPQNYL VFQLKAEDSD	600
EGHNSQLFYT ILRDP SRLFA INKESGEVFL KKQINS DHSE DLSIVVAVYD LGRPSLSTNA	660
TVKFILTDSE PSNVEVVILQ PSAEEQHQID MSIIFIAVLA GGCALLLAI FFFACTCKKK	720
AGEFKQVPEQ HGTCEERLL STPSQSVSS SLSQSESCQL SINTESENC SSSNQEQHQQ	780
TGIKHSISVP SYHTSGWHLN NCAMSIGHS HMGHISTKVQ WAKEIVTSMT VTLILVENQK	840
RRALSSQCRH KPVINTQMNQ QGSDMPITIS ATESTRVQRM GTAHCNMKRA IDCLTL	

Figure 5
SUBSTITUTE SHEET (RULE 26)

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GAATTOCCAG	AGATGAACTC	CTTGAGATTG	TTTTAAATGA	CTGCAGGTCT	GGAAGGATTG	60
CTTAAGGGTC	TCTACTTGAG	GAACCTCTAAC	AAAATTTACT	GACGTCCAGA	CCTTCTTAAG	
ACATTGCCAC	ACTGTTTCTA	GGCATGAAAA	AACTGCAAGT	TTCAACTTTG	TTTTTGGTGC	120
TGTAACGGTG	TGACAAAGAT	COGTACTTTT	TTGACGTTC	AAGTTGAAAC	AAAAACCACG	
AACTTTGATT	CTTCAAGATG	CTGCTTCTCT	TCAGAGCCAT	TCCAATGCTG	CTGTTGGGAC	180
TTGAAACTAA	GAAGTTCTAC	GACGAAGAGA	AGTCTCGGTA	AGGTTACGAC	GACAACCCTG	
TGATGGTTTT	ACAAACAGAC	TGTGAAATTG	CCCAGTACTA	CATAGATGAA	GAAGAACCCC	240
ACTACCAAAA	TGTTTGCTCG	ACACTTTAAC	GGGTCATGAT	GTATCTACTT	CTTCTTGGGG	
CTGGCACTGT	AATTGCAGTG	TTGTCACAAC	ACTCCATATT	TAACACTACA	GATATAOCTG	300
GACCGTGACA	TTAOCGTAC	AACAGTGTG	TGAGGTATAA	ATTGTGATGT	CTATATGGAC	
CAACCAATTT	COGTCTAATG	AAGCAATTTA	ATAATTCCCT	TATCGGAGTC	CGTGAGAGTG	360
GTTGGTTAAA	GGCAGATTAC	TTCGTTAAAT	TATTAAGGGA	ATAGCCTCAG	GCACTCTCAC	
ATGGGCAGCT	GAGCATCATG	GAGAGGATTG	ACCGGGAGCA	AATCTGCAGG	CAGTCCCTTC	420
TACCCGTGGA	CTCGTAGTAC	CTCTCTAAC	TGGCCCTCGT	TTAGACGTCC	GTCAGGGAAG	
ACTGCAAOCT	GGCTTTGGAT	GTGGTCAGCT	TTTCCAAAGG	ACACTTCAAG	CTTCTGAACG	480
TGACGTTGGA	CCGAAACCTA	CACCAGTCCA	AAAGGTTTCC	TGTGAAGTTC	GAAGACTTGC	
TGAAAGTGGA	GGTGAGAGAC	ATTAATGACC	ATAGCCCTCA	CTTCCCACTG	GAAATAATGC	540
ACTTTCAOCT	CCACTCTCTG	TAATTACTGG	TATCGGGAGT	GAAAGGGTCA	CTTTATTACG	
ATGTGGAGGT	GTCTGAAAGT	TCCTCTGTGG	GCACCAGGAT	TCCTTTAGAA	ATTGCAATAG	600
TACACCTCCA	CAGACTTTCA	AGGAGACACC	CGTGGTCCCTA	AGGAAATCTT	TAACGTATATC	
ATGAAGATGT	TGGGTCCAAC	TCCATCCAGA	ACTTTCAGAT	CTCAAATAAT	AGCCACTTCA	660
TACTTCTACA	ACCCAGGTTG	AGGTAGGTCT	TGAAAGTCTA	GAGTTTATTA	TCGGTGAAGT	
GCATTGATGT	GCTAACCAGA	GCAGATGGGG	TGAAATATGC	AGATTTAGTC	TTAATGAGAG	720
CGTAACTACA	CGATTGGTCT	CGTCTACCCC	ACTTTATACG	TCTAAATCAG	AATTACTCTC	
AACTGGACAG	GGAAATCCAG	CCAACATACA	TAATGGAGCT	ACTAGCAATG	GATGGGGGTG	780
TTGAOCTGTC	CCTTTAGGTC	GGTTGTATGT	ATTACCTCGA	TGATCGTTAC	CTACCCOCCAC	
TACCATCACT	ATCTGGTACT	GCAGTGGTTA	ACATCCGAGT	CCTGGACTTT	AATGATAACA	840
ATGGTAGTGA	TAGACCATGA	CGTCAOCCAT	TGTAGGCTCA	GGACCTGAAA	TTACTATTGT	
GCCAGTGTG	TGAGAGAAGC	AOCATTGCTG	TGGAOCTAGT	AGAGGATGCT	CCTCTGGGAT	900
CGGGTCACAA	ACTCTCTTCG	TGGTAACGAC	AOCTGGATCA	TCTCCTACGA	GGAGAOCTA	
ACCTTTTGT	GGAGTTACAT	GCTACTGAAG	ATGATGAAGG	AGTGAATGGA	GAAATTGTTT	960
TGGAAACAA	CCTCAATGTA	CGATGACTGC	TACTACTTCC	TCACTTACCT	CTTTAACAAA	
ATGGATTGAG	CACCTTGGCA	TCTCAAGAGG	TACGTCAGCT	ATTTAAATTT	AACTCCAGAA	1020
TACCTAAGTC	GTGAAACCGT	AGAGTTCTCC	ATGCAGTCCA	TAAATTTTAA	TTGAGGTCTT	

Figure 6A
SUBSTITUTE SHEET (RULE 26)

9/18

CTGGCAGTGT TACTCTTGAA GGCCAAAGTTG ATTTTGAGAC CAAGCAGACT TACGARTTTG	1080
GACCGTCACA ATGAGAACTT CCGGTTCAAC TAAARCTCTG GTTCGTCTGA ATGCTTAAAC	
AGGTACAAGC CCAAGATTG GGGCCCAACC CACTGACTGC TACTTGTAAG GTAACGTTC	1140
TOCATGTTG GGTCTAACC CCGGGGTTGG GTGACTGACG ATGAACATTT CATTGACAAG	
ATATACTTGA TGTAAATGAT AATACCCAG CCATCACTAT TACCCCTCTG ACTACTGTAA	1200
TATATGAAC ACATTACTA TTATGGGGTC GGTAGTGATA ATGGGGAGAC TGATGACATT	
ATGCAGGAGT TGOCTATATT CCAGAAACAG CCACAAAGGA GAACTTTATA GCTCTGATCA	1260
TACGTCTCA ACGGATATAA GGTCTTTGTC GGTGTTTCTT CTGAAATAT CGAGACTAGT	
GCACTACTGA CAGAGCCTCT GGATCTAATG GACAAGTTG CTGTACTCTT TATGGACATG	1320
CGTGATGACT GTCTCGGAGA CCTAGATTAC CTGTTCAAGC GACATGAGAA ATACCTGTAC	
AGCACTTTAA ACTACAGCA GCTTATGAGG ACAGTTACAT GATAGTTACC ACCTCTACTT	1380
TCGTGAAATT TGATGTGCTT CGAATACTCC TGTCAATGTA CTATCAATGG TGGAGATGAA	
TAGACAGGGA AAACATAGCA GCGTACTCTT TGACAGTAGT TGCAGAAGAC CTTGGCTTCC	1440
ATCTGTCCCT TTTGTATCGT CGCATGAGAA ACTGTCATCA ACGTCTTCTG GAACCGAAGG	
CCTCATTGAA GACCAAAAAG TACTACACAG TCAAGGTTAG TGATGAGAAT GACAATGCAC	1500
GGAGTAACCT CTGGTTTTTC ATGATGTGTC AGTTCCAATC ACTACTCTTA CTGTTACGTG	
CTGTATTTTC TAAACCCAG TATGAAGCTT CTATTCTGGA AAATAATGCT CCAGGCTCTT	1560
GACATAAAG ATTTGGGGTC ATACTTCGAA GATAAGACCT TTTATTACGA GGTCCGAGAA	
ATATACTAC AGTGATAGCC AGAGACTCTG ATAGTGATCA AAATGGCAAA GTAAATTACA	1620
TATATTGATG TCACTATCGG TCTCTGAGAC TATCACTAGT TTTACCGTTT CATTTAATGT	
GACTTGTGGA TGCAAAAGTG ATGGGCCAGT CACTAACAAAC ATTTGTTTCT CTTGATGCCG	1680
CTGAACACCT ACGTTTTAC TACCCGGTCA GTGATTGTTG TAAACAAAGA GAACTACGCC	
ACTCTGGAGT ATTGAGAGCT GTTAGGTCTT TAGACTATGA AAACTTAAA CAACTGGATT	1740
TGAGACCTCA TAACTCTGTA CAATCCAGAA ATCTGATACT TTTTGAATTT GTTGACCTAA	
TTGAAATTGA AGCTGCAGAC AATGGGATCC CTCAACTCTC CACTCGGTT CAACTAAATC	1800
AACTTTAACT TCGACGTCTG TTACCTAGG GAGTTGAGAG GTGAGCGCAA GTTGATTAG	
TCAGAATAGT TGATCAAAAT GATAATTGCC CTGTGATAAC TAATCCTCTT CTTAATAATG	1860
AGTCTTATCA ACTAGTTTAA CTATTAAAGG GAACTATAG ATTACGAGAA GAATTAATAG	
GCTCGGGTGA AGTTCTGCTT CCATCAGCG CTCTCAAAA CTATTAGTT TTCCAGCTCA	1920
CGAGCCCACT TCAAGACGAA GGGTAGTCGC GAGGAGTTTT GATAAATCAA AAGGTGAGT	
AAGCCGAGGA TTCAGATGAA GGGCACAAC CCAGCTGTT CTATACCATA CTGAGAGATC	1980
TTCCGCTCCT AAGTCTACTT CCGTGTGTA GGGTCGACAA GATATGGTAT GACTCTCTAG	
CAAGCAGATT GTTTGCCATT AACAAAGAAA GTGGTGAAGT GTTCTGAAA AAACAATTAA	2040
GTTOGTCTAA CAAACGGTAA TTGTTTCTTT CACCACTTCA CAAGGACTTT TTTGTTAATT	
ACTCTGACCA TTCAGAGGAC TTGAGCATAG TAGTTGCAGT GTATGACTTG GGAAGACCTT	2100
TGAGACTGGT AAGTCTCCTG AACTCGTATC ATCAACGTCA CATACTGAAC CCTTCTGGAA	
CATTATCCAC CAATGCTACA GTTAAATTCA TCCTCACCGA CTCTTTTCTT TCTAACGTTG	2160
GTAATAGGTG GTTACGATGT CAATTTAAGT AGGAGTGGCT GAGAAAAGGA AGATTGCAAC	

Figure 6B

SUBSTITUTE SHEET (RULE 26)

10/18

AAGTCGTTAT	TTTGCAACCA	TCTGCAGAAG	AGCAGCACCA	GATCGATATG	TCCATTATAT	2220
TTCAGCAATA	AAACGTTGGT	AGACGTCCTC	TOGTCGTGGT	CTAGCTATAC	AGGTAATATA	
TCATTGCAGT	GCTGGCTGGT	GGTTGTGCTT	TGCTACTTTT	GGCCATCTTT	TTTGTGGCCT	2280
AGTAAOGTCA	CGACCGACCA	CCAACACGAA	ACGATGAAAA	CCGGTAGAAA	AAACACCGGA	
GTACTTGTA	AAAGAAAGCT	GGTGAATTTA	AGCAGGTACC	TGAACAACAC	GGAACATGCA	2340
CATGAACATT	TTTCTTTCGA	CCACTTAAT	TOGTCCATGG	ACTTGTGTG	CCTGTACGT	
ATGAAGAACG	CCTGTTAAGC	ACCCATCTC	CCAGTCGGT	CTCTTCTTCT	TTGTCTCAGT	2400
TACTTCTTGC	GGACAATTCG	TGGGGTAGAG	GGTCAGCCA	GAGAAGAAGA	ARCAGAGTCA	
CTGAGTCATG	CCAACTCTCC	ATCAATACTG	AATCTGAGAA	TTGCAGCGTG	TCCTCTAACC	2460
GACTCAGTAC	GGTTGAGAGG	TAGTTATGAC	TTAGACTCTT	AACGTCGCAC	AGGAGATTGG	
AAGAGCAGCA	TCAGCAAACA	GGCATAAAGC	ACTCCATCTC	TGTACCATCT	TATCACACAT	2520
TTCTCGTCGT	AGTCGTTTGT	COGTATTTTC	TGAGGTAGAG	ACATGGTAGA	ATAGTGTGTA	
CTGTTTGGCA	CCTGGACAAT	TGTGCAATGA	GCATAAGTGG	ACATTCTCAC	ATGGGGCACA	2580
GACCAACCGT	GGACCTGTTA	ACACGTTACT	CGTATTACAC	TGTAAGAGTG	TACCCCGTGT	
TTAGTACAAA	GGTACAGTGG	GCAAAGGAGA	TAGTGACTTC	AATGACAGTG	ACTCTGATAC	2640
AATCATGTTT	CCATGTCACC	CGTTTCTCT	ATCACTGAAG	TTACTGTCAC	TGAGACTATG	
TAGTGGAGAA	TCAGAAAAGA	AGAGCATTGA	GCAGCCAATG	CAGGCACAAG	CCAGTGCTCA	2700
ATCACTCTCT	AGTCTTTTCT	TCTCGTAAC	CGTCGGTTAC	TCCCGTGTC	GGTCACGAGT	
ATACACAGAT	GAATCAGCAG	GGTTCOGACA	TGCGATAAC	TATTTACGCC	ACCGAATCAA	2760
TATGTGTCTA	CTTAGTCGTC	CCAAGGCTGT	ACGGCTATTG	ATAAAGTCGG	TGGCTTAGTT	
CAAGGGTCCA	GAAATGGGA	ACTGCACATT	GCAATATGAA	AAGGGCTATA	GACTGTCTTA	2820
GTTCOCAGGT	CTTTACCT	TGACGTGTA	OGTTATACCT	TTCCCGATAT	CTGACAGAAT	
CTCTGTAGCT	CCTGTATATT	ACAATACCTA	CCATGCAAGA	ATGCCTAACC	TGCACATACC	2880
GAGACATCGA	GGACATATA	TGTTATGGAT	GGTACGTTCT	TACGGATTGG	ACGTGTATGG	
GAAOCATACC	CTTAGAGACC	CTTATTACCA	TATCAATAAT	CCTGTTGCTA	ATCGGATGCA	2940
CTTGGTATGG	GAATCTCTGG	GAATAATGGT	ATAGTTATTA	GGACAACGAT	TAGCTACGT	
GGCGGAATAT	GAAAGAGATT	TAGTCAACAG	AAGTGCAACG	TTATCTCCGC	AGAGATCGTC	3000
CGCGCTATA	CTTTCTTAA	ATCAGTTGTC	TTACGTTGC	ATTAGAGGCG	TCTTAGCAG	
TAGCAGATAC	CAAGAATTCA	ATTACAGTCC	GCAGATATCA	AGACAGCTTC	ATCCTTCAGA	3060
ATCGTCTATG	GTCTTAAAGT	TAATGTCAGG	CGTCTATAGT	TCTGTGGAAG	TAGGAAGTCT	
AATTGCTACA	ACCTTTTAA	CATTAGGCAT	GCAAGTGAGA	ATGCACAAAG	GCAAGTGCTT	3120
TTAAGCATGT	TGGAAAATTA	GTAATCCGTA	CGTTCACTCT	TACGTGTTTC	CGTTCACGAA	
TAGCATGAAA	GCTAAATATA	TGGAGTCTCC	CCTTTCCCTC	TGATGGATGG	GGGGAGACAC	3180
ATCGTACTTT	CGATTTATAT	ACCTCAGAGG	GGAAAGGGAG	ACTACCTACC	CCCCTCTGTG	
AGGACAGTGC	ATAAATATAC	AGCTGCTTTC	TATTTGCATT	TCACTTGGGA	ATTTTTTGT	3240
TOCTGTACG	TATTTATATG	TCGACGAAAG	ATAACGTA	AGTGAACCT	TAAAAACAA	
TTTTTTACAT	ATTTATTTT	CCTGAATTGA	ATGTGACATT	GTCCTGTAC	CTAACTAGCA	3300
AAAAAATGTA	TAAATAAAAA	GGACTTAACT	TACACTGTAA	CAGGACAGTG	GATTGATCGT	

Figure 6C

SUBSTITUTE SHEET (RULE 26)

11/18

ATTAAATCCA CAGACCTACA GTCAATATT TGAGGGCCCC TGAAACAGCA CATCAGTCAG 3360
TAATTTAGGT GTCTGGATGT CAGTTTATAA ACTCCCGGGG ACTTTGTCGT GTAGTCAGTC

GACCTAAAGT GGCCTTTTTA CTTTACGAG CTCTGGGTC TGCCCTCTGT GTTAATCAGC 3420
CTGGATTTC CCGGAAAAAT GAAAATCGTC GAGGACCCAG ACGGGAGACA CAATTAGTCG

CCCTGGTCAA GTCCTGAGTA GGATCATGGC GTTTTATAT GCATCTCACC TACTTTGGAC 3480
GGGACCAGTT CAGGACTCAT CCTAGTACCG CAAAAATATA CGTAGAGTGG ATGAAACCTG

GTGATTTACA CATAATAGGA AACGCTTGGT TTCAGTGAAG TCTGTGTTGT ATATATTCTG 3540
CACTAAATGT GTATTATCCT TTGCGAACCA AAGTCACTTC AGACACAACA TATATAAGAC

TTATATACAC GCATTTTGTG TTTGTGTATA TATTTCAAGT CCATTCAGAT ATGTGTATAT 3600
AATATATGTG CGTAAAACAC AAACACATAT ATAAAGTTCA GGTAAGTCTA TACACATATA

AGTGCAGACC TTGTAAATTA AATATTCTGA TACTTTTCC TCAATAAATA TTAAAT
TCACGCTGG AACATTTAAT TTATAAGACT ATGAAAAAGG AGTTATTTAT AAATTTA

Figure 6D

SUBSTITUTE SHEET (RULE 26)

12 / 18

MVCCGPGRML LGWAGLLVLA ALCLLQVPGA QAAACEPVRI PLCKSLPWNM TKMPNHLHHS	60
TQANAILAME QFEGLLGTHC SPDLLFFLCA MYAPICTIDF QHEPIKPCKS VCERARQGCE	120
PILIKYRHSW PESLACDELP VYDRGVCISP EAIVTADGAD FPMDSSTGHC RGASSERCKC	180
KPV RATQKTY FRNNYNYVIR AKVKEVKMKC HDVTAVVEVK EILKASLVNI PRDTVNLVTT	240
SGCLCPPLTV NEEYVIMGYE DEERSRLLLV EGSIAEKWED RLGGKKVKRWD MKLRHLGLGK	300
TDASDSTQNG KSGRNSNPRP ARS.	

13 / 18

AAGCCTGGGA	CCATGGTCTG	CTGCGGCCCG	GGACGGATGC	TGCTAGGATG	GGCCGGGTG	60
TTCGGACCCT	GGTACCAGAC	GACGCCGGGC	CCTGCCTACG	ACGATCCTAC	CCGGCCCAAC	
CTAGTCTCTG	CTGCTCTCTG	CCTGCTCCAG	GTGCCCAGAG	CTCAGGCTGC	AGCCTGTGAG	120
GATCAGGACC	GACGAGAGAC	GGACGAGGTC	CACGGGCCTC	GAGTCCGACG	TCCGACACTC	
CCTGTCCGCA	TCCCCTGTGT	CAAGTCCCTT	CCTGGAACA	TGACCAAGAT	GCCCAACCAC	180
GGACAGGCGT	AGGGCGACAC	GTTCAGGGAA	GGGACCTTGT	ACTGGTTCTA	CGGGTTGGTG	
CTGCACCACA	GCACCCAGGC	TAACGCCATC	CTGGCCATGG	AACAGTTCTGA	AGGGCTGCTG	240
GACGTGGTGT	CGTGGGTCCG	ATTGCGGTAG	GACCGGTACC	TTGTCAAGCT	TCCCGACGAC	
GGCACCCACT	GCAGCCCGGA	TCTTCTCTTC	TTCCTCTGTG	CAATGTACGC	ACCCATTTCG	300
CCGTGGGTGA	CGTCGGGCCT	AGAAGAGAAG	AAGGAGACAC	GTTACATGCG	TGGGTAAACG	
ACCATCGACT	TCCAGCACGA	GCCCATCAAG	CCCTGCAAGT	CTGTGTGTGA	GCGCGCCCGA	360
TGGTAGCTGA	AGGTCGTGCT	CGGGTAGTTC	GGGACGTTCA	GACACACACT	CGCGCGGGCT	
CAGGGCTGCG	AGCCCATTCT	CATCAAGTAC	CGCCACTCGT	GGCCGAAAG	CTTGGCTGCG	420
GTCCCGACGC	TCGGGTAAGA	GTAGTTCATG	GCGGTGAGCA	CCGGCCTTTC	GAACCGGACG	
GACGAGCTGC	CGGTGTACGA	CGCGGGCGTG	TGCATCTCTC	CTGAGGCCAT	CGTCACCGCG	480
CTGCTCGACG	GCCACATGCT	GGCGCCGCAC	ACGTAGAGAG	GACTCCGGTA	GCAGTGGCGC	
GACGGAGCGG	ATTTTCCTAT	GGATTCAAGT	ACTGGACACT	GCAGAGGGGC	AAGCAGCGAA	540
CTGCCTCGCC	TAAAAGGATA	CCTAAGTTCA	TGACCTGTGA	CGTCTCCCCG	TTCGTGCTTT	
CGTTGCAAAAT	GTAAGCCTGT	CAGAGCTACA	CAGAAGACCT	ATTTCCGGAA	CAATTACAAC	600
GCAACGTTTA	CATTCCGACA	GTCTCGATGT	GTCTTCTGGA	TAAAGGCCTT	GTTAATGTGT	
TATGTCATCC	GGGCTAAAGT	TAAAGAGGTA	AAGATGAAAT	GTCATGATGT	GACCGCCGTT	660
ATACAGTAGG	CCCGATTTC	ATTCTCCAT	TTCTACTTTA	CAGTACTACA	CTGGCGGCAA	
GTGGAAGTGA	AGGAAATTCT	AAAGGCATCA	CTGGTAAACA	TTCCAAGGGA	CACCGTCAAT	720
CACCTTCACT	TCCTTTAAGA	TTTCCGTAGT	GACCATTGT	AAGGTTCCCT	GTGGCAGTTA	
CTTTATACCA	CCTCTGGCTG	CCTCTGTCCCT	CCACTTACTG	TCAATGAGGA	ATATGTCATC	780
GAAATATGGT	GGAGACCGAC	GGAGACAGGA	GGTGAATGAC	AGTTACTCCT	TATACAGTAG	
ATGGGCTATG	AAGACGAGGA	ACGTTCACGG	TTACTCTTGG	TAGAAGGCTC	TATAGCTGAG	840
TACCCGATAC	TTCTGCTCCT	TGCAAGGTCC	AATGAGAACC	ATCTTCCGAG	ATATCGACTC	
AAGTGGAAGG	ATCGGCTTGG	TAAGAAAGTC	AAGCGCTGGG	ATATGAAACT	CCGACACCTT	900
TTACCTTCC	TAGCCGAACC	ATTCTTTCAG	TTCCGACCC	TATACTTTGA	GGCTGTGGAA	
GGACTGGGTA	AAACTGATGC	TAGCGATTCC	ACTCAGAATC	AGAAGTCTGG	CAGGAACTCT	960
CCTGACCCAT	TTTGACTACG	ATCGCTAAGG	TGAGTCTTAG	TCTTCAGACC	GTCTTGAGA	

Figure 8A
SUBSTITUTE SHEET (RULE 26)

14 / 18

AATCCCCGGC	CAGCACGCAG	CTAAATCCTG	AAATGTAAAA	GGCCACACCC	ACGGACTCCC	1020
TTAGGGGCCC	GTCGTGCGTC	GATTTAGGAC	TTTACATTTT	CCGGTGTGGG	TGCCTGAGGG	
TTCTAAGACT	GGCGCTGGTG	GACTAACAAA	GGAAAACCGC	ACAGTTGTGC	TCGTGACCGA	1080
AAGATTCTGA	CCGCGACCAC	CTGATTGTTT	CCTTTTGGCG	TGTCAACACG	AGCACTGGCT	
TTGTTTACCG	CAGACACCGC	GTGGCTACCG	AAGTTACTTC	CGGTCCCCTT	TCTCCTGCTT	1140
AACAAATGGC	GTCTGTGGCG	CACCGATGGC	TTCAATGAAG	GCCAGGGGAA	AGAGGACGAA	
CTTAATGGCG	TGGGGTTAGA	TCCTTTAATA	TGTTATATAT	TCTGTTTCAT	CAATCACGTG	1200
GAATTACCGC	ACCCCAATCT	AGGAAATTAT	ACAATATATA	AGACAAAGTA	GTTAGTGCAC	
GGGACTGTTT	TTTTGCAACC	AGAATAGTAA	ATTAAATATG	TTGATGCTAA	GGTTTCTGTA	1260
CCCTGACAAG	AAAACGTTGG	TCTTATCAT	TAATTTATAC	AACTACGATT	CCAAAGACAT	
CTGGACTCCC	TGGGTTTAAT	TTGGTGTTC	GTACCTGAT	TGAGAATGCA	ATGTTTCATG	1320
GACCTGAGGG	ACCCAAATTA	AACCACAAGA	CATGGGACTA	ACTCTTACGT	TACAAAGTAC	
TAAAGAGAGA	ATCCTGGTCA	TATCTCAAGA	ACTAGATATT	GCTGTAAGAC	AGCCTCTGCT	1380
ATTTCTCTCT	TAGGACCAGT	ATAGAGTTCT	TGATCTATAA	CGACATTCTG	TCGGAGACGA	
GCTGCGCTTA	TAGTCTTGTC	TTTGATGTC	TTTGTCCATT	TCCCTCATGC	TGTGAAAGTT	1440
CGACGCGAAT	ATCAGAACAC	AAACATACGG	AAACAGGTAA	AGGGAGTACG	ACACTTTCAA	
ATACATGTTT	ATAAAGGTAG	AACGGCATTT	TGAAATCAGA	CACTGCACAA	GCAGAGTAGC	1500
TATGTACAAA	TATTTCCATC	TTGCCGTAAA	ACTTTAGTCT	GTGACGTGTT	CGTCTCATCG	
CCAACACCAG	GAAGCATTTA	TGAGGAAACG	CCACACAGCA	TGACTTATTT	TCAAGATTGG	1560
GGTTGTGGTC	CTTCGTAAAT	ACTCCTTTGC	GGTGTGTCGT	ACTGAATAAA	AGTTCTAACC	
CAGGCAGCAA	AATAAATAGT	GTTGGGAGCC	AAGAAAAGAA	TATTTTGCCT	GGTTAAGGGG	1620
GTCCGTCGTT	TTATTTATCA	CAACCTCGG	TTCTTTTCTT	ATAAAACGGA	CCAATTCCCC	
CACACTGGAA	TCAGTAGCCC	TTGAGCCATT	AACAGCAGTG	TTCTTCTGGC	AAGTTTTTGA	1680
GTGTGACCTT	AGTCATCGGG	AACTCGGTAA	TTGTGTCAC	AAGAAGACCG	TTCAAAAAC	
TTTGTTTATA	AATGTATTCA	CGAGCATTAG	AGATGAACTT	ATAACTAGAC	ATCTGTTGTT	1740
AAACAAGTAT	TTACATAAGT	GCTCGTAATC	TCTACTTGAA	TATTGATCTG	TAGACAACAA	
ATCTCTATAG	CTCTGCTTCC	TTCTAAATCA	AACCCATGTG	TGGATGCTCC	CTCTCCATTTC	1800
TAGAGATATC	GAGACGAAGG	AAGATTTAGT	TTGGGTAACA	ACCTACGAGG	GAGAGGTAAG	

Figure 8B
SUBSTITUTE SHEET (RULE 26)

15 / 18

ATAAATAAAT TTGGCTTGCT GTATTGGCCA GGAAAAGAAA GTATTAAAGT ATGCATGCAT 1860
TATTTATTTA AACCGAACGA CATAACCGGT CCTTTTCTTT CATAATTTC A TACGTACGTA

GTGCACCAGG GTGTTATTTA ACAGAGGTAT GTAACCTCTAT AAAAGACTAT AATTTACAGG 1920
CACGTGGTCC CACAATAAAT TGTCTCCATA CATTGAGATA TTTTCTGATA TTAAATGTCC

ACACGGAAAT GTGCACATTT GTTTACTTTT TTTCTTCCTT TTGCTTTGGG CTTGTGATTT 1980
TGTGCCTTTA CACGTGTAAA CAAATGAAAA AAAGAAGGAA AACGAAACCC GAACACTAAA

TGGTTTTTGG TGTGTTTATG TCTGTATTTT GGGGGGTGGG TAGGTTTAAG CCATTGCACA 2040
ACCAAAAACC ACACAAATAC AGACATAAAA CCCCCACCC ATCCAAATTC GGTAAACGTGT

TTCAAGTTGA ACTAGATTAG AGTAGACTAG GCTCATTGGC CTAGACATTA TGATTTGAAT 2100
AAGTTCAACT TGATCTAATC TCATCTGATC CGAGTAACCG GATCTGTAAT ACTAAACTTA

TTGTGTGTT TAATGCTCCA TCAAGATGTC TAATAAAAGG AATATGGTTG TCAACAGAGA 2160
AACACAACAA ATTACGAGGT AGTTCTACAG ATTATTTTCC TTATACCAAC AGTTGTCTCT

CGACAACAAC AACAAA
GCTGTTGTTG TTGTTT

16 / 18

MVCGSPGGML LLRAGLLALA ALCLLRVPGA RAAACEPVRI PLCKSLPWNM TKMPNHLHHS	60
TQANAILAIE QFEGLLGTHC SPDLLFFLCA MYAPICTIDF QHEPIKPCKS VCERARQGCE	120
PILIKYRHSW PENLACEELP VYDRGVCISP EAIVTADGAD FPMDSNGNC RGASSERCKC	180
KPIRATQKTY FRNNYNYVIR AKVKEIKTKC HDVTAVVEVK EILKSSLVNI PRDTVNLVTS	240
SGCLCPPLNV NEEYIIMGYE DEERSRLLLV EGSIAEKWD RLGKKVKRWD MKLRHLGLSK	300
SDSSNSDSTQ SQKSGRNSNP RQARN.	

Figure 9

SUBSTITUTE SHEET (RULE 26)

17 / 18

GGCGGAGCGG	GCCTTTTGGC	GTCCACTGCG	CGGCTGCACC	CTGCCCCATC	TGCCGGGATC	60
CCGCCTCGCC	CGGAAAACCG	CAGGTGACGC	GCCGACGTGG	GACGGGGTAG	ACGGCCCTAG	
ATGGTCTGCG	GCAGCCCGGG	AGGGATGCTG	CTGCTGCGGG	CCGGGCTGCT	TGCCCTGGCT	120
TACCAGACGC	CGTCGGGCCC	TCCCTACGAC	GACGACGCCC	GGCCCGACGA	ACGGGACCGA	
GCTCTCTGCC	TGCTCCGGGT	GCCCCGGGCT	CGGGCTGCAG	CCTGTGAGCC	CGTCCGCATC	180
CGAGAGACGG	ACGAGGCCCA	CGGGCCCCGA	GCCCGACGTC	GGACACTCGG	GCAGGCGTAG	
CCCCTGTGCA	AGTCCCTGCC	CTGGAACATG	ACTAAGATGC	CCAACCACCT	GCACCACAGC	240
GGGACACAGT	TCAGGGACGG	GACCTGTGAC	TGATTCTACG	GGTTGGTGGA	CGTGGTGTCTG	
ACTCAGGCCA	ACGCCATCCT	GGCCATCGAG	CAGTTCGAAG	GTCTGCTGGG	CACCCACTGC	300
TGAGTCCGGT	TGCGGTAGGA	CCGGTAGCTC	GTCAAGCTTC	CAGACGACCC	GTGGGTGACG	
AGCCCCGATC	TGCTCTTCTT	CCTCTGTGCC	ATGTACGCGC	CCATCTGCAC	CATTGACTTC	360
TCGGGGCTAG	ACGAGAAGAA	GGAGACACGG	TACATGCGCG	GGTAGACGTG	GTAACGTGAAG	
CAGCACGAGC	CCATCAAGCC	CTGTAAGTCT	TGTGTGCGAGC	GGGCCCCGCA	GGGCTGTGAG	420
GTCGTGCTCG	GGTAGTTCCG	GACATTGAGA	CACACGCTCG	CCCGGGCCGT	CCCGACACTC	
CCCATACTCA	TCAAGTACCG	CCACTCGTGG	CCGGAGAACC	TGGCCTGCGA	GGAGCTGCCA	480
GGGTATGAGT	AGTTCATGGC	GGTGAGCACC	GGCCTCTTGG	ACCGGACGCT	CCTCGACGGT	
GTGTACGACA	GGGGCGTGTG	CATCTCTCCC	GAGGCCATCG	TTACTGCGGA	CGGAGCTGAT	540
CACATGCTGT	CCCCGCACAC	GTAGAGAGGG	CTCCGGTAGC	AATGACGCCT	GCCTCGACTA	
TTTCCTATGG	ATTCTAGTAA	CGGAACTGT	AGAGGGGCAA	GCAGTGAACG	CTGTAAATGT	600
AAAGGATACC	TAAGATCATT	GCCTTTGACA	TCTCCCCGTT	CGTCACTTGC	GACATTTACA	
AAGCCTATTA	GAGCTACACA	GAAGACCTAT	TTCCGGAACA	ATTACAACTA	TGTCATTCCG	660
TTCCGATAAT	CTCGATGTGT	CTTCTGGATA	AAGGCCTTGT	TAATGTTGAT	ACAGTAAGCC	
GCTAAAGTTA	AAGAGATAAA	GACTAAGTGC	CATGATGTGA	CTGCAGTAGT	GGAGGTGAAG	720
CGATTTCAAT	TTCTCTATTT	CTGATTACAG	GTACTACACT	GACGTCATCA	CCTCCACTTC	
GAGATTCTAA	AGTCCTCTCT	GGTAAACATT	CCACGGGACA	CTGTCAACCT	CTATACCAGC	780
CTCTAAGATT	TCAGGAGAGA	CCATTTGTAA	GGTGCCCTGT	GACAGTTGGA	GATATGGTCG	
TCTGGCTGCC	TCTGCCCTCC	ACTTAATGTT	AATGAGGAAT	ATATCATCAT	GGGCTATGAA	840
AGACCGACGG	AGACGGGAGG	TGAATTACAA	TTACTCCTTA	TATAGTAGTA	CCCGATACTT	

Figure 10A
SUBSTITUTE SHEET (RULE 26)

18 / 18

GATGAGGAAC	GTTCCAGATT	ACTCTTGGTG	GAAGGCTCTA	TAGCTGAGAA	GTGGAAGGAT	900
CTACTCCTTG	CAAGGTCTAA	TGAGAACCAC	CTTCCGAGAT	ATCGACTCTT	CACCTTCCTA	
CGACTCGGTA	AAAAAGTTAA	GCGCTGGGAT	ATGAAGCTTC	GTCATCTTGG	ACTCAGTAAA	960
GCTGAGCCAT	TTTTTCAATT	CGCGACCCTA	TACTTCGAAG	CAGTAGAACC	TGAGTCATTT	
AGTGATTCTA	GCAATAGTGA	TTCCACTCAG	AGTCAGAAGT	CTGGCAGGAA	CTCGAACCCC	1020
TCACTAAGAT	CGTTATCACT	AAGGTGAGTC	TCAGTCTTCA	GACCGTCCTT	GAGCTTGGGG	
CGGCAAGCAC	GCAACTAAAT	CCCGAAATAC	AAAAAGTAAC	ACAGTGGACT	TCCTATTAAG	1080
GCCGTTCGTG	CGTTGATTTA	GGGCTTTATG	TTTTTCATTG	TGTCACCTGA	AGGATAATTC	
ACTTACTTGC	ATTGCTGGAC	TAGCAAAGGA	AAATTGCACT	ATTGCACATC	ATATTCTATT	1140
TGAATGAACG	TAACGACCTG	ATCGTTTCCT	TTTAACGTGA	TAACGTGTAG	TATAAGATAA	
GTTTACTATA	AAAATCATGT	GATAACTGAT	TATTACTTCT	GTTTCTCTTT	TGGTTTCTGC	1200
CAATGATAT	TTTTAGTACA	CTATTGACTA	ATAATGAAGA	CAAAGAGAAA	ACCAAAGACG	
TTCTCTCTTC	TCTCAACCCC	TTTGTAAATG	TTTGGGGGCA	GACTCTTAAG	TATATTGTGA	1260
AAGAGAGAAG	AGAGTTGGGG	AAACATTACC	AAACCCCCGT	CTGAGAATTC	ATATAACACT	
GTTTCTATT	TCACTAATCA	TGAGAAAAAC	TGTTCTTTTG	CAATAATAAT	AAATTAAACA	1320
CAAAAGATAA	AGTGATTAGT	ACTCTTTTGT	ACAAGAAAAAC	GTTATTATTA	TTTAATTTGT	
TGCTGTTACC	AGAGCCTCTT	TGCTGAGTCT	CCAGATGTTA	ATTTACTTTT	TGCACCCCAA	1380
ACGACAATGG	TCTCGGAGAA	ACGACTCAGA	GGTCTACAAT	TAAATGAAAG	ACGTGGGGTT	
TTGGAATGC	AATATTGGAT	GAAAAGAGAG	GTTTCTGGTA	TTACAGAAAA	GCTAGATATG	1440
AACCTTACG	TTATAACCTA	CTTTTCTCTC	CAAAGACCAT	AAGTGTCTTT	CGATCTATAC	
CCTTAAACA	TACTCTGCCG	ATCTAATTAC	AGCCTTATTT	TTGTATGCCT	TTTGGGCATT	1500
GGAATTTTGT	ATGAGACGGC	TAGATTAAATG	TCGGAATAAA	AACATACGGA	AAACCCGTAA	
CTCCTCATGC	TTAGAAAGTT	CCAAATGTTT	ATAAAGGTAA	AATGGCAGTT	TGAAGTCAAA	1560
GAGGAGTACG	AATCTTTCAA	GGTTTACAAA	TATTTCCATT	TTACCGTCAA	ACTTCAGTTT	
TGTCACATAG	GCAAAGCAAT	CAAGCACCAG	GAAGTGTTTA	TGAGGAAACA	ACACCCAAGA	1620
ACAGTCTATC	CGTTTGGTTA	GTTCTGGTGC	CTTCAAAAT	ACTCTTTTGT	TGTGGGTCT	
TGAATTATTT	TTGAGACTGT	CAGGAAGTAA	AATAAATAGG	AGCTTAAGAA	AGAACATTTT	1680
ACTTAATAAA	AACTCTGACA	GTCCTTCATT	TTATTATATC	TCGAATTCCT	TCTTGTAATA	
GCCTGATTGA	GAAGCACAAC	TGAAACCAGT	AGCCGCTGGG	GTGTTAATGG	TAGCATTCCT	1740
CGGACTAACT	CTTCGTGTTG	ACTTTGGTCA	TCGGCGACCC	CACAATTACC	ATCGTAAGAA	
CTTTTGCCAA	TACATTTGAT	TTGTTTCATG	ATATATTAAT	CAGCATTAGA	GAAATGAATT	1800
GAAACCGTT	ATGTAAACTA	AACAAGTACT	TATATAATTA	GTCGTAACTT	CTTTACTTAA	
ATAACTAGAC	ATCTGCTGTT	ATCACCATAG	TTTTGTTTAA	TTTGCTTCCT	TTTAAATAAA	1860
TATTGATCTG	TAGACGACAA	TAGTGGTATC	AAAACAAATT	AAACGAAGGA	AAATTTATTT	
CCCATTGGTG	AAAGTCAAAA	AAAAAAAAAA	AAA			
GGGTAACCAC	TTTCAGTTTT	TTTTTTTTTT	TTT			

Figure 10B
SUBSTITUTE SHEET (RULE 26)

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/10942

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.

US CL : 530/300, 350; 514/2; 536/23.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/300, 350; 514/2; 536/23.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

DIALOG (MEDLINE, BIOSIS, EMBASE, WPI, USPATFULL) AUTHOR AND WORD. search terms: e.g. cerberus, xenopus

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y, P	BOUWMEESTER et al. Cerberus is a head-inducing secreted factor expressed in the anterior endoderm of Spemann's organizer. Nature. 15 August 1996, Vol. 382, No. 6592, pages 595-601, see entire document.	1-15

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Z" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

29 AUGUST 1997

Date of mailing of the international search report

11 SEP 1997

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/10942

A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

A01N 37/18; A61K 38/00; C07K 1/00, 2/00, 4/00, 7/00, 14/00, 16/00, 17/00; C07H 21/02, 21/04